

Pathophysiology of metabolic dysfunction-associated steatotic liver disease

Rocío Gallego-Durán^{1,2,3}, Paloma Carrillo-Fernández^{1,2}, and Jordi Gracia-Sancho^{3,4*}

¹UCM Digestive Diseases, Hospital Universitario Virgen del Rocío, SeLiver Group, Instituto de Biomedicina de Sevilla (HUVR/CSIC/US), Seville; ²Department of Medicine, Universidad de Sevilla, Seville; ³Centro de Investigación Biomédica en Red en Enfermedades Hepáticas y Digestivas (CIBEREHD), Madrid; ⁴Hepatic Vascular Biology Laboratory, IDIBAPS – Hospital Clínic de Barcelona, Barcelona. Spain

Abstract

Metabolic dysfunction-associated steatotic liver disease (MASLD) is the most prevalent liver pathology worldwide, affecting more than one-third of the general population. Its pathophysiology is complex and multifactorial, involving metabolic, immunological, genetic, and environmental alterations that interact synergistically to determine the patient's phenotype. This review provides an in-depth analysis of the main mechanisms involved in the development and progression of MASLD, including energy excess and *de novo* lipogenesis, insulin resistance, lipid metabolism disturbances, oxidative stress and mitochondrial dysfunction, inflammation mediated by the inflammasome, genetic factors, and intestinal dysbiosis with increased gut permeability. These alterations converge in hepatic microcirculatory dysfunction, which promotes hepatic stellate cell activation and progression toward fibrosis, advanced chronic liver disease, and hepatocellular carcinoma. Moreover, MASLD is associated with a significant increase in cardiovascular, metabolic, and neoplastic risk, reinforcing its multisystemic nature and the need for a comprehensive clinical approach. In this context, personalized medicine emerges as a key tool for risk stratification and therapeutic optimization, using biomarkers, genetic profiling, and microbiome analysis. A detailed understanding of the pathophysiological mechanisms of MASLD is essential for the development of effective preventive and therapeutic strategies, and for improving the prognosis of patients affected by this silent yet clinically, economically, and socially impactful disease.

Keywords: Steatosis. Liver cells. Disease mechanisms.

Fisiopatología de la enfermedad hepática asociada a disfunción metabólica

Resumen

La enfermedad hepática asociada a disfunción metabólica (MASLD, Metabolic dysfunction Associated Steatotic Liver Disease) constituye la patología hepática de mayor prevalencia en todo el mundo, afectando a más de un tercio de la población general. Su fisiopatología es compleja y multifactorial, pues involucra alteraciones metabólicas, inmunitarias, genéticas y ambientales que interactúan de manera sinérgica para determinar el fenotipo del paciente. Este trabajo revisa en profundidad los principales mecanismos implicados en el desarrollo y la progresión de la MASLD, incluyendo el exceso de energía y la lipogénesis *de novo*, la resistencia a la insulina, las alteraciones en el metabolismo lipídico, el estrés oxidativo y la disfunción mitocondrial, la inflamación mediada por el inflammasoma, los factores genéticos y la disbiosis intestinal con aumento de la permeabilidad intestinal. Estas alteraciones convergen en una disfunción microcirculatoria hepática que favorece la activación

*Correspondence:

Jordi Gracia-Sancho
E-mail: jordi.graciasancho@gmail.com

Date of reception: 01-10-2025

Date of acceptance: 05-11-2025

DOI: 10.24875/CGME.M25000039

Available online: 17-03-2026

Clín. Gastroenterol. Méx. (Eng). 2025;1(4):313-322

www.clinicastroenterologiademexico.com

de las células hepáticas estrelladas y la progresión hacia fibrosis, enfermedad hepática crónica avanzada y carcinoma hepatocelular. Además, esta enfermedad se asocia con un incremento significativo del riesgo cardiovascular, metabólico y neoplásico, lo que refuerza su carácter multisistémico y la necesidad de un abordaje clínico integral. En este contexto, la medicina personalizada emerge como una herramienta clave para la estratificación del riesgo y la optimización terapéutica, mediante el uso de biomarcadores, perfiles genéticos y análisis del microbioma. La comprensión detallada de los mecanismos fisiopatológicos de la MASLD es esencial para el desarrollo de estrategias preventivas y terapéuticas eficaces, y para mejorar el pronóstico de los pacientes afectados por esta enfermedad silenciosa, pero de gran impacto clínico, económico y social.

Palabras clave: Esteatosis. Células hepáticas. Mecanismos de enfermedad.

Introduction

Metabolic dysfunction-associated steatotic liver disease (MASLD) has consolidated as the main cause of chronic liver disease globally, with an estimated prevalence of around 30-40% of the adult population and an increasing trend in recent decades¹. In 2023, an international multidisciplinary consensus proposed a nomenclature change from NAFLD/NASH (non-alcoholic fatty liver disease/non-alcoholic steatohepatitis) to MASLD/MASH (metabolic dysfunction-associated steatotic liver disease/metabolic dysfunction-associated steatohepatitis), framing it under the umbrella of SLD (steatotic liver disease). MASLD was defined by the presence of steatosis ($\geq 5\%$ of hepatocytes by imaging or histology) along with one or more cardiometabolic risk factors and absence of significant alcohol intake². This update, already incorporated in the 2024 clinical guideline of the European Association for the Study of the Liver (EASL), the European Association for the Study of Diabetes (EASD), and the European Association for the Study of Obesity (EASO)³, reinforces the emphasis on metabolic dysfunction as the pathogenic axis and standardizes the criteria for its screening, risk stratification, and management.

The etiology of MASLD is heterogeneous and converges in the interaction of lifestyle factors (hypercaloric diet, sedentary lifestyle), obesity, and metabolic syndrome, with relevant contributions from genetic and epigenetic factors, and intestinal dysbiosis⁴⁻¹⁰. In addition, exposure to environmental pollutants has been increasingly linked to the development of steatosis and disease progression, underscoring its multifactorial nature¹¹⁻¹³. Therefore, MASLD is recognized today as a multisystemic disease that transcends the liver.

From a pathophysiological standpoint, the evolution of MASLD is best explained by the “multiple parallel hits” hypothesis, which replaces the old “two-hit” model¹⁴. It converges, synchronously, insulin resistance, lipotoxicity, and dysfunction of the gut-pancreas-liver axis, generating a hepatic microenvironment prone to

chronic low-grade inflammation and fibrosis. Insulin resistance (hepatic and systemic) drives *de novo* lipogenesis, increased flux of free fatty acids to the liver, and accumulation of lipotoxic species (ceramides, diacylglycerols), with compensatory hyperinsulinemia and inter-organ crosstalk (adipose tissue, muscle, pancreas) that perpetuates the metabolic vicious cycle^{15,16}.

Oxidative stress and mitochondrial dysfunction constitute central nodes of progression: substrate excess and lipid overload alter mitochondrial function, favor lipid peroxidation, endoplasmic reticulum stress, and generation of reactive oxygen species (ROS), with hepatocellular damage and increased profibrotic signaling pathways¹⁷⁻¹⁹. Such processes are amplified by damage-associated molecular pattern (DAMP) and pathogen-associated molecular pattern (PAMP) signals, and by metabolites derived from the intestinal microbiota (lipopolysaccharides, endogenous ethanol, and short-chain fatty acids), which, when crossing a more permeable intestinal barrier, activate innate and adaptive pathways in the liver^{10,20,21}. All this contributes to the transition toward steatohepatitis (MASH), defined histologically by steatosis with ballooning, lobular inflammation, and fibrosis (Fig. 1).

The persistence of metabolic-inflammatory injury precipitates hepatocyte death (through various pathways, including apoptosis, necrosis, and ferroptosis), sinusoidal endothelial cell dysfunction, and recruitment of the immune compartment (macrophages/Kupffer cells, monocytes, lymphocytes), generating a proinflammatory microenvironment, marked by the release of proinflammatory cytokines (tumor necrosis factor alpha [TNF α], interleukin [IL] 1 β , transforming growth factor beta [TGF β], among others) that will activate hepatic stellate cells^{22,23}. These cells undergo a myofibroblastic phenotypic transition, with proliferative, contractile, and migratory properties, and secrete excessive extracellular matrix, driving fibrogenesis^{24,25}. In the “omics” era, at the single-cell level, subpopulations of activated hepatic stellate cells with distinct fibrogenic and proinflammatory signatures have been

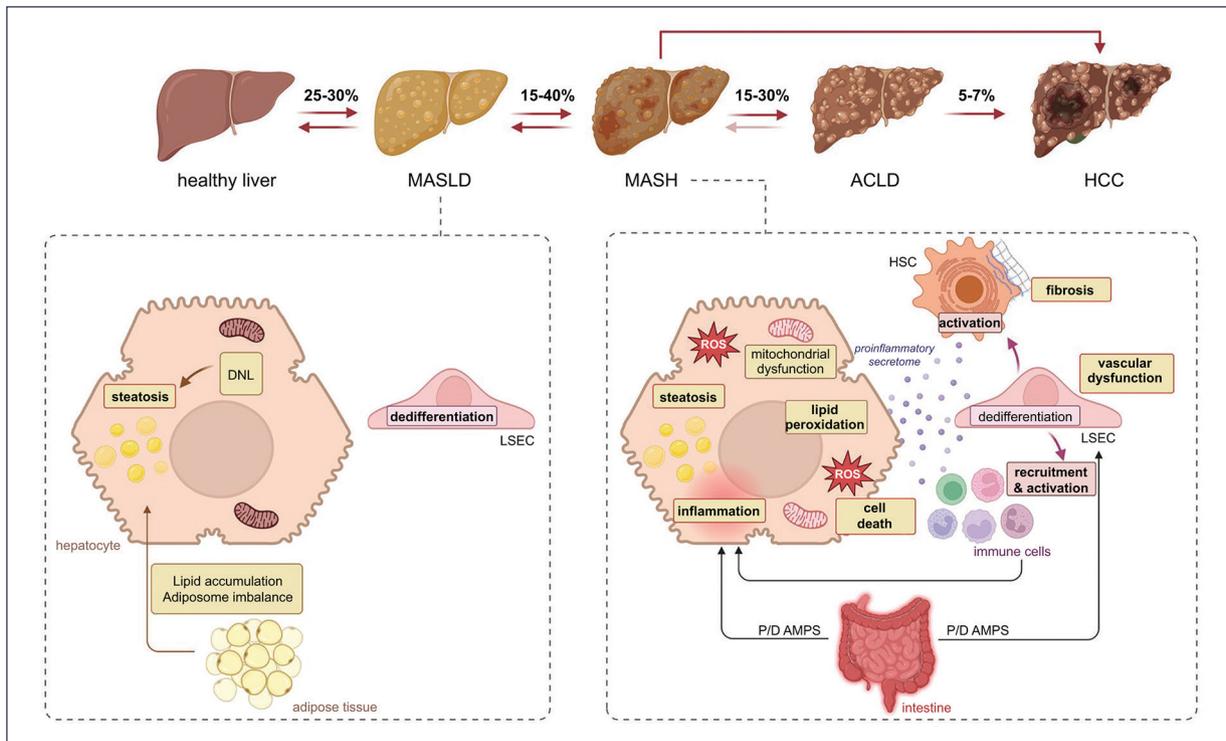


Figure 1. Pathophysiology of liver disease associated with metabolic dysfunction. This figure shows the complex mechanisms that lead to progression from metabolic dysfunction-associated steatotic liver disease (MASLD) to metabolic dysfunction-associated steatohepatitis (MASH), and beyond. Among the main contributing factors are dyslipidemia, endothelial dysfunction, and chronic inflammation, which lead to the accumulation of lipids in the hepatocyte. In more advanced stages, mitochondrial dysfunction plays a key role in the progression of the disease. Additionally, the figure highlights the role of dysregulation of the intestine-liver axis and inflammation of adipose tissue in disease progression. Furthermore, fibrogenesis – driven by the activation of hepatic stellate cells and the deposition of extracellular matrix – is shown as a distinctive hallmark of the transition to more advanced liver disease. This multifactorial process culminates in liver injury, inflammation, and fibrosis characteristic of MASH. ACLD: advanced chronic liver disease; DNL: de novo lipogenesis; LSEC: liver sinusoidal endothelial cell; HSC: hepatic stellate cell; P/D AMPS: pathogen/danger-associated molecular patterns; ROS: reactive oxygen species.

described, revealing the functional heterogeneity of this cell type and potentially facilitating the development of specific therapeutic targets^{26,27}.

Clinically, the degree of fibrosis is the main predictor of hepatic and extrahepatic events and mortality, above other histological features, which would justify non-invasive fibrosis stratification strategies and prioritization of antifibrotic objectives²⁸⁻³⁰. Fibrosis can progress to cirrhosis, with architectural distortion and regenerative nodules that would compromise hepatic function, and in a fraction of patients, can progress to hepatocellular carcinoma (HCC). It should be noted that the MASLD-MASH-cirrhosis-HCC continuum is not always linear: a non-negligible proportion of HCC in MASLD occurs in patients without established cirrhosis, although with advanced fibrosis in many cases, which would complicate screening and early detection³¹.

Beyond the liver, MASLD significantly increases cardiovascular risk, being the main cause of death in these patients, and is associated with higher all-cause mortality, as well as an increase in extrahepatic neoplasms and cardiometabolic comorbidity^{32,33}. This systemic risk profile reinforces the need for integrated management strategies with other clinical specialties, focused on lifestyle change, control of risk factors, and, when appropriate, specific therapies, in line with current guidelines.

Overall, MASLD is a complex and multifactorial disorder in which dysregulated lipogenesis and lipid metabolism, insulin resistance, oxidative and endoplasmic reticulum stress, immune-mediated inflammation, genetic/epigenetic susceptibility, and intestinal dysbiosis converge. The following sections of this review will address in depth the main mechanistic axes with the

objective of integrating basic, translational, and clinical evidence that supports the pathophysiology and progression of the disease.

Most relevant pathophysiological aspects of MASLD

Energy excess and de novo lipogenesis

Energy excess, especially derived from the consumption of hypercaloric diets rich in simple sugars (such as fructose) and saturated fats, constitutes one of the main triggering factors of MASLD. When energy intake exceeds the metabolic needs of the organism, the liver acts as a storage organ, accumulating triglycerides in hepatocytes. This process is exacerbated by the activation of *de novo* lipogenesis, a metabolic pathway that converts carbohydrates into fatty acids.

De novo lipogenesis is regulated by transcription factors, such as SREBP-1c (sterol regulatory element-binding protein 1c) and ChREBP (carbohydrate response element-binding protein), which are activated in response to the presence of elevated levels of insulin and glucose. These factors induce the expression of key enzymes, such as acetyl coenzyme A carboxylase and fatty acid synthase, responsible for fatty acid synthesis. These fatty acids are subsequently esterified with glycerol to form triglycerides, which accumulate in the cytoplasm of hepatocytes³⁴⁻³⁶.

In addition, energy excess inhibits fatty acid oxidation and the secretion of triglyceride-rich lipoproteins (VLDL, very low density lipoproteins), further contributing to intrahepatic lipid accumulation^{37,38}. This lipid overload not only generates hepatic steatosis but also induces lipotoxicity, a phenomenon in which certain lipids, such as ceramides and diacylglycerols, alter cellular function and generate oxidative stress, inflammation, and apoptosis³⁹. Therefore, this imbalance between intake and energy expenditure and the liver's capacity to handle lipids represents the first step in the pathogenic cascade of MASLD, setting the stage for progression toward more severe forms, such as MASH and hepatic fibrosis.

Insulin resistance

Insulin resistance is one of the most relevant pathophysiological pillars in the development of MASLD. Under normal conditions, insulin regulates energy metabolism by promoting glucose uptake in peripheral tissues, inhibiting lipolysis in adipose tissue, and favoring glycogen synthesis in the liver. However, in situations of insulin resistance, frequently associated with

visceral obesity, sedentary lifestyle, and hypercaloric diet, these functions are compromised.

In adipose tissue, insulin resistance causes uncontrolled activation of lipolysis, which generates an increase in the release of free fatty acids into the portal circulation⁴⁰. These free fatty acids are taken up by the liver, where they are re-esterified into triglycerides, contributing to hepatic steatosis. In addition, free fatty acids can activate proinflammatory signaling pathways, such as that of nuclear factor kappa B and JNK (Jun-N-terminal kinase), which interfere with the insulin signaling pathway, perpetuating the state of resistance^{41,42}.

In the liver, insulin resistance alters the balance between gluconeogenesis and lipogenesis^{43,44}. Although insulin loses its capacity to inhibit hepatic glucose production, it maintains its lipogenic effect, resulting in a metabolic paradox, since glucose production increases while lipids accumulate. This situation favors metabolic dysfunction and the development of lipotoxicity.

At the molecular level, a decrease in the phosphorylation of the insulin receptor and key proteins such as IRS-1 and Akt has been described, which compromises signal transduction⁴⁵⁻⁴⁷. In addition, chronic low-grade inflammation, mediated by cytokines such as TNF- α and IL-6, contributes to its inhibition⁴⁸. Overall, insulin resistance not only promotes hepatic fat accumulation but also establishes a proinflammatory and prooxidative environment that facilitates progression toward MASH.

Lipid metabolism disturbances

Lipid metabolism in the liver is a dynamic process that involves the uptake, synthesis, oxidation, and export of fatty acids. In the context of MASLD, this balance is profoundly altered, favoring lipid accumulation in hepatocytes and contributing to the onset and progression of the disease.

Under physiological conditions, the liver takes up fatty acids from the diet, from adipose tissue lipolysis, and from *de novo* lipogenesis⁴⁹. These fatty acids can be oxidized in the mitochondria, peroxisomes, or endoplasmic reticulum, or be esterified to form triglycerides that are stored or exported as VLDL⁵⁰.

In MASLD, this process becomes unbalanced by several mechanisms. First, fatty acid uptake increases due to insulin resistance and increased peripheral lipolysis, which elevates the lipid load in the liver⁴⁹. Second, fatty acid oxidation is compromised by mitochondrial dysfunction, which reduces the capacity to eliminate excess lipids by the hepatocyte⁵¹. Finally, triglyceride

export as VLDL becomes insufficient, either due to defects in apolipoprotein synthesis or secretion system overload^{52,53}.

Overall, this imbalance favors the accumulation of toxic lipids, such as ceramides, diacylglycerols, and free cholesterol, inducing lipotoxicity. These lipids alter various cellular signaling pathways, induce oxidative stress and inflammation, and promote hepatocyte apoptosis. In addition, the accumulation of free cholesterol in cell membranes can destabilize organelles, especially mitochondria and the endoplasmic reticulum. Therefore, the alteration of lipid metabolism not only contributes to hepatic steatosis but also acts as a driver of progression toward advanced stages of the disease.

Oxidative stress and mitochondrial dysfunction

As previously mentioned, oxidative stress and mitochondrial dysfunction are key elements in the progression of MASLD from simple steatosis to MASH and hepatic fibrosis. Excessive accumulation of lipids in hepatocytes, especially free fatty acids and cholesterol, overloads the mitochondria, which are responsible for the β -oxidation of fatty acids. This overload generates an increase in ROS production, exceeding cellular antioxidant capacity⁵⁴⁻⁵⁷.

ROS, such as hydrogen peroxide (H_2O_2), superoxide anion (O_2^-), and hydroxyl radicals ($OH\cdot$), can damage lipids, proteins, and nucleic acids, altering the integrity of cellular and organellar membranes⁵⁸. In the liver, this damage translates into lipid peroxidation, endoplasmic reticulum dysfunction, activation of inflammatory pathways, and cell death by apoptosis or necrosis⁵⁹.

Mitochondrial dysfunction also involves an alteration in mitochondrial dynamics (fusion and fission), loss of membrane potential, decreased oxidative phosphorylation, and release of cytochrome c, which activates the apoptotic cascade. In addition, damaged mitochondria release danger signals, such as DAMPs, which activate the innate immune system, including the NLRP3 inflammasome⁶⁰.

Another relevant aspect is the decreased activity of antioxidant enzymes, such as superoxide dismutase, catalase, and glutathione peroxidase, which aggravates the redox imbalance⁶¹. Oxidative stress can also interfere with the function of nuclear receptors, among them PPAR- α (peroxisome proliferator-activated receptor alpha) and FXR (farnesoid x receptor), which regulate lipid metabolism and inflammation⁶².

Therefore, oxidative stress and mitochondrial dysfunction not only contribute to direct hepatocellular

damage, but also amplify the inflammatory and fibrogenic response, and are central elements in the transition from MASLD toward more severe stages.

Inflammation and inflammasome activation

Inflammation is a central component in the progression of MASLD, as the liver becomes an immunologically active organ, where multiple signals of cellular and metabolic damage trigger both innate and adaptive inflammatory responses. One of the most relevant mechanisms is inflammasome activation, especially the NLRP3 complex, which acts as a sensor of cellular stress⁶³⁻⁶⁵.

The NLRP3 inflammasome is activated in response to danger signals (DAMPs and PAMPs), such as bacterial endotoxins, cholesterol crystals, saturated fatty acids, and extracellular adenosine triphosphate. Its activation leads to caspase-1 activation, which in turn processes the proforms of IL-1 β and IL-18 into their active forms. These cytokines are potent inflammatory mediators that amplify hepatic necroinflammation, favoring the recruitment of immune cells such as neutrophils, monocytes, and lymphocytes^{63,64,66,67}.

In addition, inflammation in MASLD is modulated by the polarization of hepatic macrophages (also known as Kupffer cells) toward a proinflammatory phenotype (M1), which secretes TNF- α , IL-6, and other cytokines that interfere with insulin signaling and promote hepatocellular damage^{68,69}. This chronic low-grade inflammation also affects sinusoidal endothelial cells and hepatic stellate cells, facilitating their activation and contributing to fibrogenesis⁷⁰.

Inflammasome activation, in addition to having local implications in the liver, can also generate systemic effects, such as insulin resistance, endothelial dysfunction, and alterations in other organs^{71,72}, so that the combination of inflammation and inflammasome activation represents a bridge between altered metabolism and the immune response, being fundamental mechanisms in the progression of MASLD.

Genetic factors

Genetic factors play a fundamental role in individual susceptibility to the development and progression of disease⁷³. Although environmental and metabolic factors are key determinants, genetic variability explains why some individuals develop severe forms of disease even in the absence of metabolic risk factors, while

others present mild steatosis without disease progression despite having multiple risk factors.

Among the most studied genes is *PNPLA3* (patatin-like phospholipase domain-containing protein 3), whose polymorphism rs738409 (I148M) is strongly associated with hepatic fat accumulation, inflammation, and fibrosis, as well as an increased risk of developing cirrhosis and HCC. This mutant allele reduces the lipase activity of the protein, favoring triglyceride retention in hepatocytes. Its effect is independent of insulin resistance, making it a robust genetic risk marker^{74,75}.

Another relevant gene is *TM6SF2* (transmembrane 6 superfamily member 2), whose polymorphism rs58542926 (E167K) is associated with a decrease in VLDL secretion, which favors the accumulation of intrahepatic lipids⁷⁶. However, although this gene appears to protect against cardiovascular diseases by reducing plasma lipid levels, it has been shown to increase the risk of hepatic fibrosis⁷⁷.

The gene *MBOAT7* (membrane-bound o-acyltransferase domain-containing 7) has also been implicated in the regulation of phospholipid metabolism. Its variant rs641738 is associated with inflammation and fibrosis, possibly by altering the composition of cell membranes and intracellular signaling⁷⁸.

More recently, the gene *HSD17B13*, variant rs72613567, has been identified; its loss of function appears to have a protective effect in progression toward MASH and fibrosis⁷⁹⁻⁸¹. This finding has opened new therapeutic avenues based on genetic modulation.

Genetic factors not only modulate the clinical and phenotypic expression of MASLD, but also offer opportunities for personalized medicine, allowing identification of high-risk individuals and development of targeted therapeutic strategies.

Dysbiosis and intestinal permeability

The gut-liver axis plays a crucial role in the pathophysiology of liver diseases, specifically MASLD, and its alteration by intestinal dysbiosis and increased intestinal barrier permeability contributes significantly to progression⁸². The intestine harbors a diverse microbial community that participates in digestion, nutrient metabolism, immune modulation, and epithelial barrier protection⁸³. In MASLD, this microbiota is altered both in its composition and function, a phenomenon known as dysbiosis.

Intestinal dysbiosis is characterized by a decrease in beneficial bacteria (such as *Faecalibacterium prausnitzii* and *Akkermansia muciniphila*) and an increase in

proinflammatory species (such as *Enterobacteriaceae* and *Proteobacteria*)^{84,85}. This alteration favors the production of harmful metabolites, such as endogenous ethanol, and the imbalance in the proportion of short-chain fatty acids, ammonia, and lipopolysaccharides, which can cross the intestinal barrier when it is compromised, a phenomenon known as leaky gut⁸⁶.

Increased intestinal permeability allows the passage of endotoxins and other bacterial products into the portal circulation, reaching the liver and activating receptors such as TLR4 (toll-like receptor 4) in Kupffer cells and hepatocytes. This activation triggers inflammatory responses, cytokine production, and inflammasome activation, contributing to hepatic necroinflammation and progression toward MASH⁸⁷.

In addition, dysbiosis can influence bile acid metabolism, altering the signaling of receptors such as FXR and TGR5 (Takeda G protein-coupled receptor 5), which regulate energy homeostasis, inflammation, and insulin sensitivity⁸⁸. It has also been observed that certain microbial profiles can modulate the expression of hepatic genes related to lipogenesis and fibrosis, highlighting the crucial role of intestinal dysbiosis in disease progression⁸⁹.

Hepatic microcirculatory dysfunction and fibrosis

Hepatic microcirculatory dysfunction represents a direct consequence of the interaction of the pathophysiological mechanisms described in MASLD. The liver possesses a unique vascular architecture, composed of sinusoids that allow efficient exchange of nutrients, metabolites, and immune cells between portal blood and hepatocytes. In MASLD, this microcirculation is progressively altered, contributing to the functional deterioration of the organ and the development of fibrosis.

Lipid accumulation in hepatocytes, oxidative stress, and chronic inflammation induce endothelial damage in sinusoidal endothelial cells. These cells lose their specialized phenotype, characterized by fenestrations and low expression of adhesion molecules, and adopt a proinflammatory, vasoconstrictive, and procoagulant state⁹⁰. This transformation increases hepatic vascular tone and reduces exchange between blood and hepatocytes, promoting local hypoxia and metabolic dysfunction. In turn, hypoxia activates factors, such as HIF-1 α (hypoxia-inducible factor 1-alpha), which promote pathological angiogenesis and hepatic stellate cell activation. These cells, in response to inflammatory cytokines (TGF- β , platelet-derived growth factor, IL-1 β), differentiate

into myofibroblasts, acquiring contractile, migratory, and secretory properties. The result is excessive production of extracellular matrix components (collagen type I and III, fibronectin), which are deposited in the space of Disse, further altering microcirculation^{91,92}.

Hepatic fibrosis is characterized by an imbalance between the synthesis and degradation of extracellular matrix, due to the inhibition of metalloproteinases and activation of their inhibitors⁹³. This process distorts hepatic architecture, compromises portal blood flow, and can evolve toward cirrhosis, with regenerative nodule formation and loss of hepatic function. Fibrosis, combined with the increase in vascular tone derived from sinusoidal endothelial cell dedifferentiation and hepatic stellate cell activation, leads to an increase in intrahepatic vascular resistance, a primary factor in the development of portal hypertension⁹⁴.

Conclusions

MASLD represents a clinical and scientific challenge of growing relevance, given its high global prevalence and its multisystemic character. Throughout this review, its dynamic and multifactorial character has been analyzed, being the result of a complex interaction of genetic, environmental, metabolic, and cellular factors that converge in a cascade of pathological events.

Moreover, the pathophysiological progression of MASLD from hepatic steatosis to advanced fibrosis has significant clinical implications that affect not only the liver but also the general state of the patient, whose clinical consequences derive from the interaction of metabolic, inflammatory, genetic, immunological, and microcirculatory alterations⁹⁵.

Systemically, it is associated with a significantly increased risk of cardiovascular disease, which represents the leading cause of death in these patients. Insulin resistance, dyslipidemia, chronic inflammation, and endothelial dysfunction contribute to the development of atherosclerosis, hypertension, and major cardiovascular events⁹⁶⁻⁹⁹. Furthermore, a higher incidence of type 2 diabetes mellitus, metabolic syndrome, and certain types of extrahepatic cancers, such as colorectal and breast cancer, has been observed¹⁰⁰⁻¹⁰³. Intestinal dysbiosis and systemic inflammation may also affect renal function, the central nervous system, and overall immune status^{82,104,105}.

In the liver, fat accumulation and progressive cellular damage can evolve toward MASH, characterized by necroinflammation, hepatocellular ballooning, and hepatic stellate cell activation^{25,106}. This stage can lead

toward the development of fibrosis, which can advance toward hepatic cirrhosis, with distortion of hepatic architecture, portal hypertension, hepatic insufficiency, and HCC risk^{92,107}. It is important to highlight that this progression is not always linear, and some patients can develop HCC in the absence of cirrhosis, that is, in early stages of advanced chronic liver disease.

Moreover, this pathology is associated with a significant increase in cardiovascular, metabolic, and neoplastic risk, which reinforces the need for comprehensive diagnostic and therapeutic approaches. The identification of specific biomarkers, the study of genetic variants, and the analysis of the intestinal microbiome open new avenues for personalized medicine and prevention of complications. In this complex scenario, personalized medicine emerges as a promising strategy to improve the clinical management of the disease. The identification of genetic variants associated with greater susceptibility, together with the phenotypic analysis of the patient, allows a more precise risk stratification and a more rational selection of therapeutic interventions. In addition, the development of non-invasive biomarkers and molecular diagnostic tools will facilitate dynamic disease monitoring and evaluation of treatment response. Overall, personalized medicine not only optimizes clinical efficacy, but also reduces the risk of adverse effects and improves patients' quality of life, positioning itself as a fundamental pillar in the future of MASLD treatment¹⁰⁸.

Finally, from the clinical point of view, these consequences require a multidisciplinary approach in the management of the patient with MASLD, which will include hepatology, endocrinology, cardiology, and nutrition. Early identification of at-risk patients, the use of non-invasive biomarkers, and the implementation of personalized therapeutic strategies are essential to prevent complications and improve prognosis, reducing their impact on the economic burden of public health systems.

Funding

The authors acknowledge continued funding from the Instituto de Salud Carlos III (ISCIII; currently PI23/00945, DTS22/00010, DTS24/00035, and AC24/00124 to J. Gracia-Sancho; CD21/00095, PI22/01342, and PI24/02008 to R. Gallego-Durán, all co-financed by the European Union), AGAUR-Generalitat de Catalunya (2021 SGR 01322, 2021 PROD 00036, 2025 PROD 00184, and 2025 INNOV 00043) to J. Gracia-Sancho, and CIBEREHD (funded by the Instituto de Salud Carlos III).

Conflicts of interest

The authors declare no conflicts of interest.

Ethical considerations

Protection of human and animal subjects. The authors declare that no experiments were performed on humans or animals for this research.

Confidentiality, informed consent, and ethical approval. The study does not involve personal patient data nor requires ethical approval. SAGER guidelines do not apply.

Declaration on the use of artificial intelligence. The authors declare that they did not use any type of generative artificial intelligence for the writing of this manuscript.

References

1. Younossi ZM, Kalligeros M, Henry L. Epidemiology of metabolic dysfunction-associated steatotic liver disease. *Clin Mol Hepatol.* 2025;31:32-50.
2. Rinella ME, Lazarus JV, Ratziu V, Francque SM, Sanyal AJ, Kanwal F, et al. A multisociety Delphi consensus statement on new fatty liver disease nomenclature. *J Hepatol.* 2023;79:1542-56.
3. European Association for the Study of the Liver (EASL); European Association for the Study of Diabetes (EASD); European Association for the Study of Obesity (EASO). EASL-EASD-EASO Clinical Practice Guidelines. EASL-EASD-EASO Clinical Practice Guidelines on the management of metabolic dysfunction-associated steatotic liver disease (MASLD). *J Hepatol.* 2024;81:492-542.
4. Neuschwander-Tetri BA, Clark JM, Bass NM, Van Natta ML, Unalp-Arida A, Tonascia J, et al. Clinical, laboratory and histological associations in adults with nonalcoholic fatty liver disease. *Hepatology.* 2010;52:913-24.
5. Samuel VT, Shulman GI. Nonalcoholic fatty liver disease as a nexus of metabolic and hepatic diseases. *Cell Metab.* 2018;27:22-41.
6. Trépo E, Valenti L. Update on NAFLD genetics: from new variants to the clinic. *J Hepatol.* 2020;72:1196-209.
7. Loomba R, Friedman SL, Shulman GI. Mechanisms and disease consequences of nonalcoholic fatty liver disease. *Cell.* 2021;184:2537-64.
8. Cantero I, Abete I, Babio N, Arós F, Corella D, Estruch R, et al. Dietary Inflammatory Index and liver status in subjects with different adiposity levels within the PREDIMED trial. *Clin Nutr.* 2018;37:1736-43.
9. Bowden Davies KA, Sprung VS, Norman JA, Thompson A, Mitchell KL, Harrold JA, et al. Physical activity and sedentary time: association with metabolic health and liver fat. *Med Sci Sports Exerc.* 2019;51:1169-77.
10. Bahitham W, Banoun Y, Aljahdali M, Almuaiqly G, Bahshwan SM, Aljahdali L, et al. 'Trust your gut': exploring the connection between gut microbiome dysbiosis and the advancement of Metabolic Associated Steatosis Liver Disease (MASLD)/Metabolic Associated Steatohepatitis (MASH): a systematic review of animal and human studies. *Front Nutr.* 2025;12:1637071.
11. An G, Song J, Ying W, Lim W. Overview of the hazardous impacts of metabolism-disrupting chemicals on the progression of fatty liver diseases. *Mol Cell Toxicol.* 2025;21:387-97.
12. Choi MA, Rose S, Langouët S. Per- and polyfluoroalkyl substances as potentiators of hepatotoxicity in an exposome framework: current challenges of environmental toxicology. *Toxicology.* 2025;515:154167.
13. Riechelmann-Casarin L, Valente LC, Otton R, Barbisan LF, Romualdo GR. Are glyphosate or glyphosate-based herbicides linked to metabolic dysfunction-associated steatotic liver disease (MASLD)? The weight of current evidence. *Environ Toxicol Pharmacol.* 2025;116:104705.
14. Buzzetti E, Pinzani M, Tsochatzis EA. The multiple-hit pathogenesis of non-alcoholic fatty liver disease (NAFLD). *Metabolism.* 2016;65:1038-48.
15. Eslam M, Newsome PN, Sarin SK, Anstee QM, Targher G, Romero-Gómez M, et al. A new definition for metabolic dysfunction-associated fatty liver disease: an international expert consensus statement. *J Hepatol.* 2020;73:202-9.
16. Friedman SL, Neuschwander-Tetri BA, Rinella M, Sanyal AJ. Mechanisms of NAFLD development and therapeutic strategies. *Nat Med.* 2018;24:908-22.
17. Sunny NE, Bril F, Cusi K. Mitochondrial adaptation in nonalcoholic fatty liver disease: novel mechanisms and treatment strategies. *Trends Endocrinol Metab.* 2017;28:250-60.
18. Li X, Chen W, Jia Z, Xiao Y, Shi A, Ma X. Mitochondrial dysfunction as a pathogenesis and therapeutic strategy for metabolic-dysfunction-associated steatotic liver disease. *Int J Mol Sci.* 2025;26:4256.
19. Le Guillou D, Siao K, Her CL, Duwaerts CC, Maher JJ. iPSC-derived hepatocytes from patients with MASLD exhibit early mitochondrial dysfunction. *bioRxiv [Preprint].* 2025 Aug 30:2025.08.27.672733. doi: 10.1101/2025.08.27.672733.
20. Dua A, Kumari R, Singh M, Kumar R, Pradeep S, Ojesina AI, et al. Metabolic Dysfunction-Associated Steatotic Liver Disease (MASLD): the interplay of gut microbiome, insulin resistance, and diabetes. *Front Med (Lausanne).* 2025;12:1618275.
21. Che Z, Xue W, Zhao X, Hu C, Tian Y. Regulatory role and biomarker potential of gut microbiota metabolites in the progression of metabolic dysfunction-associated steatotic liver disease (MASLD) to hepatocellular carcinoma (HCC). *Clin Transl Gastroenterol.* 2025 Sep 5. doi: 10.14309/ctg.0000000000000914. Online ahead of print.
22. Malhi H, Guicciardi ME, Gores GJ. Hepatocyte death: a clear and present danger. *Physiol Rev.* 2010;90:1165-94.
23. Krenkel O, Tacke F. Liver macrophages in tissue homeostasis and disease. *Nat Rev Immunol.* 2017;17:306-21.
24. Tsuchida T, Friedman SL. Mechanisms of hepatic stellate cell activation. *Nat Rev Gastroenterol Hepatol.* 2017;14:397-411.
25. Friedman SL. Hepatic stellate cells: protean, multifunctional, and enigmatic cells of the liver. *Physiol Rev.* 2008;88:125-72.
26. Cheng S, Zou Y, Zhang M, Bai S, Tao K, Wu J, et al. Single-cell RNA sequencing reveals the heterogeneity and intercellular communication of hepatic stellate cells and macrophages during liver fibrosis. *MedComm (2020).* 2023;4:e378.
27. Ramachandran P, Dobie R, Wilson-Kanamori JR, Dora EF, Henderson BEP, Luu NT, et al. Resolving the fibrotic niche of human liver cirrhosis at single-cell level. *Nature.* 2019;575:512-8.
28. Angulo P, Kleiner DE, Dam-Larsen S, Adams LA, Bjornsson ES, Charatchoenwithaya P, et al. Liver fibrosis, but no other histologic features, is associated with long-term outcomes of patients with nonalcoholic fatty liver disease. *Gastroenterology.* 2015;149:389-97.e10.
29. Dulai PS, Singh S, Patel J, Soni M, Prokop LJ, Younossi Z, et al. Increased risk of mortality by fibrosis stage in nonalcoholic fatty liver disease: systematic review and meta-analysis. *Hepatology.* 2017;65:1557-65.
30. Younossi ZM, Loomba R, Rinella ME, Bugianesi E, Marchesini G, Neuschwander-Tetri BA, et al. Current and future therapeutic regimens for nonalcoholic fatty liver disease and nonalcoholic steatohepatitis. *Hepatology.* 2018;68:361-71.
31. Younossi ZM, Henry L. Epidemiology of non-alcoholic fatty liver disease and hepatocellular carcinoma. *JHEP Rep.* 2021;3:100305.
32. Celsa C, Pennisi G, Tulone A, Ciancimino G, Vaccaro M, Pecorella F, et al. Risk of hepatic and extrahepatic outcomes associated with metabolic dysfunction-associated steatotic liver disease and metabolic dysfunction and alcohol-associated steatotic liver disease: a systematic review and meta-analysis. *Lancet Gastroenterol Hepatol.* 2025;10:998-1012.
33. Sohrabi M, Mosalli M, Hassanzadeh P, Bahrami S, Khoonsari M, Ajdarkosh H, et al. Extrahepatic comorbidities associated with metabolic dysfunction-associated steatotic liver disease; a tertiary hospital experience. *Middle East J Dig Dis.* 2025;17:96-104.
34. Sanders FWB, Griffin JL. De novo lipogenesis in the liver in health and disease: more than just a shunting yard for glucose. *Biol Rev Camb Philos Soc.* 2016;91:452-68.
35. Denechaud PD, Dentin R, Girard J, Postic C. Role of ChREBP in hepatic steatosis and insulin resistance. *FEBS Lett.* 2008;582:68-73.
36. Shimano H, Sato R. SREBP-regulated lipid metabolism: convergent physiology - divergent pathophysiology. *Nat Rev Endocrinol.* 2017;13:710-30.
37. Ipsen DH, Lykkesfeldt J, Tveden-Nyborg P. Molecular mechanisms of hepatic lipid accumulation in non-alcoholic fatty liver disease. *Cell Mol Life Sci.* 2018;75:3313-27.
38. Puri P, Baillie RA, Wiest MM, Mirshahi F, Choudhury J, Cheung O, et al. A lipidomic analysis of nonalcoholic fatty liver disease. *Hepatology.* 2007;46:1081-90.
39. Iturbe-Rey S, Maccali C, Arrese M, Aspichueta P, Oliveira CP, Castro RE, et al. Lipotoxicity-driven metabolic dysfunction-associated steatotic liver disease (MASLD). *Atherosclerosis.* 2025;400:119053.
40. Nielsen S, Guo ZK, Johnson CM, Hensrud DD, Jensen MD. Splanchnic lipolysis in human obesity. *J Clin Invest.* 2004;113:1582-8.
41. Luukkonen PK, Zhou Y, Sädevirta S, Leivonen M, Arola J, Orešič M, et al. Hepatic ceramides dissociate steatosis and insulin resistance in patients with non-alcoholic fatty liver disease. *J Hepatol.* 2016;64:1167-75.
42. Solinas G, Naugler W, Galimi F, Lee MS, Karin M. Saturated fatty acids inhibit induction of insulin gene transcription by JNK-mediated phosphorylation of insulin-receptor substrates. *Proc Natl Acad Sci U S A.* 2006;103:16454-9.

43. Brown MS, Goldstein JL. Selective versus total insulin resistance: a pathogenic paradox. *Cell Metab.* 2008;7:95-6.
44. Bo T, Gao L, Yao Z, Shao S, Wang X, Proud CG, et al. Hepatic selective insulin resistance at the intersection of insulin signaling and metabolic dysfunction-associated steatotic liver disease. *Cell Metab.* 2024;36:947-68.
45. Hotamisligil GS, Murray DL, Choy LN, Spiegelman BM. Tumor necrosis factor alpha inhibits signaling from the insulin receptor. *Proc Natl Acad Sci U S A.* 1994;91:4854-8.
46. Aguirre V, Uchida T, Yenush L, Davis R, White MF. The c-Jun NH(2)-terminal kinase promotes insulin resistance during association with insulin receptor substrate-1 and phosphorylation of Ser(307). *J Biol Chem.* 2000;275:9047-54.
47. Gual P, Grémeaux T, Gonzalez T, Le Marchand-Brustel Y, Tanti JF. MAP kinases and mTOR mediate insulin-induced phosphorylation of Insulin Receptor Substrate-1 on serine residues 307, 612 and 632. *Diabetologia.* 2003;46:1532-42.
48. Kern PA, Ranganathan S, Li C, Wood L, Ranganathan G. Adipose tissue tumor necrosis factor and interleukin-6 expression in human obesity and insulin resistance. *Am J Physiol Endocrinol Metab.* 2001;280:E745-51.
49. Donnelly KL, Smith CI, Schwarzenberg SJ, Jessurun J, Boldt MD, Parks EJ. Sources of fatty acids stored in liver and secreted via lipoproteins in patients with nonalcoholic fatty liver disease. *J Clin Invest.* 2005;115:1343-51.
50. Rui L. Energy metabolism in the liver. *Compr Physiol.* 2014;4:177-97.
51. Sunny NE, Parks EJ, Browning JD, Burgess SC. Excessive hepatic mitochondrial TCA cycle and gluconeogenesis in humans with nonalcoholic fatty liver disease. *Cell Metab.* 2011;14:804-10.
52. Sanyal AJ, Campbell-Sargent C, Mirshahi F, Rizzo WB, Contos MJ, Sterling RK, et al. Nonalcoholic steatohepatitis: association of insulin resistance and mitochondrial abnormalities. *Gastroenterology.* 2001;120:1183-92.
53. Yamaguchi K, Yang L, McCall S, Huang J, Xing XY, Pandey SK, et al. Inhibiting triglyceride synthesis improves hepatic steatosis but exacerbates liver damage and fibrosis in obese mice with nonalcoholic steatohepatitis. *Hepatology.* 2007;45:1366-74.
54. Begrich K, Igoudjil A, Pessayre D, Fromenty B. Mitochondrial dysfunction in NASH: causes, consequences and possible means to prevent it. *Mitochondrion.* 2006;6:1-28.
55. Shi S, Wang L, Van der Laan LJW, Pan Q, Versteeg MMA. Mitochondrial dysfunction and oxidative stress in liver transplantation and underlying diseases: new insights and therapeutics. *Transplantation.* 2021;105:2362.
56. Pessayre D, Mansouri A, Haouzi D, Fromenty B. Hepatotoxicity due to mitochondrial dysfunction. *Cell Biol Toxicol.* 1999;15:367-73.
57. Sanyal AJ, Campbell-Sargent C, Mirshahi F, Rizzo WB, Contos MJ, Sterling RK, et al. Nonalcoholic steatohepatitis: association of insulin resistance and mitochondrial abnormalities. *Gastroenterology.* 2001;120:1183-92.
58. Dröge W. Free radicals in the physiological control of cell function. *Physiol Rev.* 2002;82:47-95.
59. Chen Z, Liu X, Ye Y. Oxidative stress causes liver injury via the damage of redox system and the apoptosis induced by mitochondria of broiler. *Ital J Anim Sci.* 2024;23:1469-78.
60. Zong Y, Li H, Liao P, Chen L, Pan Y, Zheng Y, et al. Mitochondrial dysfunction: mechanisms and advances in therapy. *Signal Transduct Target Ther.* 2024;9:1-29.
61. Svobodová G, Horní M, Velecká E, Boušová I. Metabolic dysfunction-associated steatotic liver disease-induced changes in the antioxidant system: a review. *Arch Toxicol.* 2025;99:1-22.
62. Zhou S, You H, Qiu S, Yu D, Bai Y, He J, et al. A new perspective on NAFLD: focusing on the crosstalk between peroxisome proliferator-activated receptor alpha (PPAR α) and farnesoid X receptor (FXR). *Biomed Pharmacother.* 2022;154:113577.
63. Mridha AR, Wree A, Robertson AAB, Yeh MM, Johnson CD, Van Rooyen DM, et al. NLRP3 inflammasome blockade reduces liver inflammation and fibrosis in experimental NASH in mice. *J Hepatol.* 2017;66:1037-46.
64. Wree A, McGeough MD, Peña CA, Schlattjan M, Li H, Inzaugarat ME, et al. NLRP3 inflammasome activation is required for fibrosis development in NAFLD. *J Mol Med (Berl).* 2014;92:1069-82.
65. Gallego-Durán R, Montero-Vallejo R, Maya-Miles D, Lucena A, Martín F, Ampuero J, et al. Analysis of common pathways and markers from non-alcoholic fatty liver disease to immune-mediated diseases. *Front Immunol.* 2021;12:667354.
66. Craven RR, Gao X, Allen IC, Gris D, Wardenburg JB, McElvania-TeKippe E, et al. *Staphylococcus aureus* alpha-hemolysin activates the NLRP3-inflammasome in human and mouse monocytic cells. *PLoS One.* 2009;4:e7446.
67. Patel S. Inflammasomes, the cardinal pathology mediators are activated by pathogens, allergens and mutagens: a critical review with focus on NLRP3. *Biomed Pharmacother.* 2017;92:819-25.
68. Tacke F, Zimmermann HW. Macrophage heterogeneity in liver injury and fibrosis. *J Hepatol.* 2014;60:1090-6.
69. Stienstra R, Saudale F, Duval C, Keshkar S, Groener JEM, Van Rooijen N, et al. Kupffer cells promote hepatic steatosis via interleukin-1 beta-dependent suppression of peroxisome proliferator-activated receptor alpha activity. *Hepatology.* 2010;51:511-22.
70. Marrone G, Shah VH, Gracia-Sancho J. Sinusoidal communication in liver fibrosis and regeneration. *J Hepatol.* 2016;65:608-17.
71. Yao J, Sterling K, Wang Z, Zhang Y, Song W. The role of inflammasomes in human diseases and their potential as therapeutic targets. *Signal Transduct Target Ther.* 2024;9:1-30.
72. Vandanmagsar B, Youm YH, Ravussin A, Galgani JE, Stadler K, Mynatt RL, et al. The NLRP3 inflammasome instigates obesity-induced inflammation and insulin resistance. *Nat Med.* 2011;17:179-89.
73. Kocas-Kilicarslan ZN, Cetin Z, Faccioli LAP, Motomura T, Amirneni S, Diaz-Aragón R, et al. Polymorphisms associated with metabolic dysfunction-associated steatotic liver disease influence the progression of end-stage liver disease. *Gastro Hep Adv.* 2023;3:67.
74. Sookoian S, Pirola CJ. Meta-analysis of the influence of I148M variant of patatin-like phospholipase domain containing 3 gene (PNPLA3) on the susceptibility and histological severity of nonalcoholic fatty liver disease. *Hepatology.* 2011;53:1883-94.
75. Romeo S, Kozlitina J, Xing C, Pertsemlidis A, Cox D, Pennacchio LA, et al. Genetic variation in PNPLA3 confers susceptibility to nonalcoholic fatty liver disease. *Nat Genet.* 2008;40:1461-5.
76. Kozlitina J, Smagris E, Stender S, Nordestgaard BG, Zhou HH, Tybjaerg-Hansen A, et al. Exome-wide association study identifies a TM6SF2 variant that confers susceptibility to nonalcoholic fatty liver disease. *Nat Genet.* 2014;46:352-6.
77. Liu YL, Reeves HL, Burt AD, Tiniakos D, McPherson S, Leathart JBS, et al. TM6SF2 rs58542926 influences hepatic fibrosis progression in patients with non-alcoholic fatty liver disease. *Nat Commun.* 2014; 5:4309.
78. Mancina RM, Dongiovanni P, Petta S, Pingitore P, Meroni M, Rametta R, et al. The MBOAT7-TMC4 variant rs641738 increases risk of nonalcoholic fatty liver disease in individuals of European descent. *Gastroenterology.* 2016;150:1219-30.e6.
79. Su W, Mao Z, Liu Y, Zhang X, Zhang W, Gustafsson JA, et al. Role of HSD17B13 in the liver physiology and pathophysiology. *Mol Cell Endocrinol.* 2019;489:119-25.
80. Abul-Husn NS, Cheng X, Li AH, Xin Y, Schurmann C, Stevis P, et al. A protein-truncating HSD17B13 variant and protection from chronic liver disease. *N Engl J Med.* 2018;378:1096-106.
81. Gil-Gómez A, Rojas A, García-Lozano MR, Muñoz-Hernández R, Gallego-Durán R, Maya-Miles D, et al. Impact of a loss-of-function variant in HSD17B13 on hepatic decompensation and mortality in cirrhotic patients. *Int J Mol Sci.* 2022;23:11840.
82. Tripathi A, Debelius J, Brenner DA, Karin M, Loomba R, Schnabl B, et al. The gut-liver axis and the intersection with the microbiome. *Nat Rev Gastroenterol Hepatol.* 2018;15:397-411.
83. Boursier J, Mueller O, Barret M, Machado M, Fizanne L, Araujo-Pérez F, et al. The severity of nonalcoholic fatty liver disease is associated with gut dysbiosis and shift in the metabolic function of the gut microbiota. *Hepatology.* 2016;63:764-75.
84. Shen F, Zheng RD, Sun XQ, Ding WJ, Wang XY, Fan JG. Gut microbiota dysbiosis in patients with non-alcoholic fatty liver disease. *Hepatobiliary Pancreat Dis Int.* 2017;16:375-81.
85. Wang J, Qin J, Li Y, Cai Z, Li S, Zhu J, et al. A metagenome-wide association study of gut microbiota in type 2 diabetes. *Nature.* 2012;490:55-60.
86. Miele L, Valenza V, La Torre G, Montalto M, Cammarota G, Ricci R, et al. Increased intestinal permeability and tight junction alterations in nonalcoholic fatty liver disease. *Hepatology.* 2009;49:1877-87.
87. Csak T, Ganz M, Pespisa J, Kodys K, Dolganiuc A, Szabo G. Fatty acid and endotoxin activate inflammasomes in mouse hepatocytes that release danger signals to stimulate immune cells. *Hepatology.* 2011;54:133-44.
88. Jiao N, Baker SS, Chapa-Rodriguez A, Liu W, Nugent CA, Tsompana M, et al. Suppressed hepatic bile acid signalling despite elevated production of primary and secondary bile acids in NAFLD. *Gut.* 2018;67: 1881-91.
89. Aron-Wisniewsky J, Vigliotti C, Witjes J, Le P, Holleboom AG, Verheij J, et al. Gut microbiota and human NAFLD: disentangling microbial signatures from metabolic disorders. *Nat Rev Gastroenterol Hepatol.* 2020;17:279-97.
90. Gracia-Sancho J, Caparrós E, Fernández-Iglesias A, Francés R. Role of liver sinusoidal endothelial cells in liver diseases. *Nat Rev Gastroenterol Hepatol.* 2021;18:411-31.
91. Copple BL. Hypoxia stimulates hepatocyte epithelial to mesenchymal transition by hypoxia-inducible factor and transforming growth factor-beta-dependent mechanisms. *Liver Int.* 2010;30:669-82.
92. Battaller R, Brenner DA. Liver fibrosis. *J Clin Invest.* 2005;115:209-18.
93. Arthur MJP. Fibrogenesis II. Metalloproteinases and their inhibitors in liver fibrosis. *Am J Physiol Gastrointest Liver Physiol.* 2000;279:G245-9.
94. Gracia-Sancho J, Marrone G, Fernández-Iglesias A. Hepatic microcirculation and mechanisms of portal hypertension. *Nat Rev Gastroenterol Hepatol.* 2019;16:221-34.

95. Younossi ZM, Koenig AB, Abdelatif D, Fazel Y, Henry L, Wymer M. Global epidemiology of nonalcoholic fatty liver disease—meta-analytic assessment of prevalence, incidence, and outcomes. *Hepatology*. 2016;64:73-84.
96. Golabi P, Owringi S, Younossi ZM. Global perspective on nonalcoholic fatty liver disease and nonalcoholic steatohepatitis—prevalence, clinical impact, economic implications and management strategies. *Aliment Pharmacol Ther*. 2024;59:S1-9.
97. Lonardo A, Ballestri S, Guaraldi G, Nascimbeni F, Romagnoli D, Zona S, et al. Fatty liver is associated with an increased risk of diabetes and cardiovascular disease—evidence from three different disease models: NAFLD, HCV and HIV. *World J Gastroenterol*. 2016;22:9674-93.
98. Mensah GA, Arnold N, Prabhu SD, Ridker PM, Welty FK. Inflammation and Cardiovascular Disease: 2025 ACC Scientific Statement: A Report of the American College of Cardiology. *J Am Coll Cardiol*. 2025 Sep 29:S0735-1097(25)07555-2. doi: 10.1016/j.jacc.2025.08.047. Online ahead of print.
99. Wang X, He B. Endothelial dysfunction: molecular mechanisms and clinical implications. *MedComm (2020)*. 2024;5:e651.
100. Mantovani A, Scorletti E, Mosca A, Alisi A, Byrne CD, Targher G. Complications, morbidity and mortality of nonalcoholic fatty liver disease. *Metabolism*. 2020;111:154170.
101. Ran D, Xin CL, Ma Y, Lu Y. Increased risk of colorectal adenomas with metabolic-associated fatty liver disease components. *Clin Res Hepatol Gastroenterol*. 2024;48:102302.
102. Mantovani A, Petracca G, Beatrice G, Csermely A, Tilg H, Byrne CD, et al. Non-alcoholic fatty liver disease and increased risk of incident extrahepatic cancers: a meta-analysis of observational cohort studies. *Gut*. 2022;71:778-88.
103. Ampuero J, Aller R, Gallego-Durán R, Banales JM, Crespo J, García-Monzón C, et al. The effects of metabolic status on non-alcoholic fatty liver disease-related outcomes, beyond the presence of obesity. *Aliment Pharmacol Ther*. 2018;48:1260-70.
104. Musso G, Gambino R, Tabibian JH, Ekstedt M, Kechagias S, Hamaguchi M, et al. Association of non-alcoholic fatty liver disease with chronic kidney disease: a systematic review and meta-analysis. *PLoS Med*. 2014;11:e1001680.
105. Mikkelsen ACD, Kjærgaard K, Schapira AHV, Mookerjee RP, Thomsen KL. The liver-brain axis in metabolic dysfunction-associated steatotic liver disease. *Lancet Gastroenterol Hepatol*. 2025;10:248-58.
106. Brunt EM, Wong VWS, Nobili V, Day CP, Sookoian S, Maher JJ, et al. Nonalcoholic fatty liver disease. *Nat Rev Dis Primers*. 2015;1:15080.
107. Schuppan D, Surabattula R, Wang XY. Determinants of fibrosis progression and regression in NASH. *J Hepatol*. 2018;68:238-50.
108. Priego-Parra BA, Gallego-Durán R, Román-Calleja BM, Velarde-Ruiz Velasco JA, Romero-Gómez M, Gracia-Sancho J. Advancing precision medicine in metabolic dysfunction-associated steatotic liver disease. *Trends Endocrinol Metab*. 2025;36:1000-13.